

Contents lists available at ScienceDirect

Agriculture, Ecosystems and Environment

journal homepage: www.elsevier.com/locate/agee



Vegetation cover and food availability shapes the foraging activity of rodent pests in and around maize fields

Christopher I. Imakando^{a,b,*}, G. Mandela Fernández-Grandon^a, Grant R. Singleton^a, Steven R. Belmain^a

^a Natural Resources Institute, University of Greenwich, Chatham Maritime, Kent ME4 4TB, UK

^b Department of Zoology and Aquatic Sciences, Copperbelt University, P.O Box 21692, Kitwe, Zambia

ARTICLE INFO

Keywords: Rodent pests Mastomys natalensis Foraging activity Maize field Giving-up-density Landscape of fear

ABSTRACT

Evidence-based information for smallholder farmers on where and when to conduct rodent management is vital given that most are resource poor and depend on agriculture for food and income. However, there is scarce information on how the foraging activity of rodent pests changes over agricultural cropping seasons. We used the concepts of giving-up-density (GUD) and landscape of fear to monitor how the foraging activity of rodent pests changes in and around maize (*Zea mays*) fields over the cropping season. We tested the hypothesis that the foraging activity of rodent pests will be influenced by vegetation cover, perceived predation risk and food availability. *Mastomys natalensis* was the dominant species in all maize fields (n = 3, 87.05 % of the total captures). We observed that the foraging activity of rodent activity in the natural habitat and along the border was higher than inside the maize fields. During the germination stage, rodent activity in and around the maize fields. During the harvest stage, the foraging activity was higher in the maize fields than along the border and in the natural habitat. These results can be used to guide smallholder farmers where and when to focus rodent control measures during different stages of the cropping season. An additional approach would be to develop strategies that could potentially increase rodent fear perceptions in cropping landscapes.

1. Introduction

Rodent pests damage maize crops before and after harvest (Skonhoft et al., 2006; Swanepoel et al., 2017). Mostly, rodent pests damage maize crops during the germination (Mulungu et al., 2005) and maturation stages (Mulungu, 2017). At the germination stage, rodents dig up and consume germinating maize seeds (Mulungu et al., 2007) leading to either a regular distribution of damage in mosaic fields or a more random distribution in monoculture fields (Mulungu et al., 2005). At the maturation stage, rodents consume both fresh and dry grains when the maize plants are standing or on the ground (Mulungu, 2017). Population dynamics and competition for available food resources partly accounts for observed heterogeneous damage patterns (Mohr et al., 2003). Predation risk, land preparation methods and soil type can also account for the heterogenous damage by indirectly or directly affecting the population dynamics of rodents (Mulungu et al., 2005). Therefore, either random or stratified sampling methods could be used to assess rodent damage in maize fields (Mulungu et al., 2007). In maize cropping systems, little is known both about how the foraging activity of rodent pests changes over the cropping season and how this may affect crop damage, particularly in relation to harbourage provided by field margin vegetation and the maize crop itself. Maintaining field margin vegetation is increasingly recognised as important in facilitating crop pollination and conservation biological control of insect pests (Arnold et al., 2021; Ochieng et al., 2022), but such vegetation could potentially exacerbate the presence of rodent pests (Jacob, 2008; Rodríguez-Pastor et al., 2016).

Understanding the foraging activity of rodent pests is important to enable sustainable control to reduce their impact and damage (Belmain, 2010; Krijger et al., 2017). Foraging activity has been strongly correlated with vegetation cover in several studies. In Philippines, *Rattus tanezumi* spent more time foraging at the centre of the rice fields than on the field edges (border), where there was less vegetation cover (Jones et al., 2017). Evidence suggests that rodents in agricultural landscapes

* Corresponding author at: Natural Resources Institute, University of Greenwich, Chatham Maritime, Kent ME4 4TB, UK. *E-mail address:* imakandochristopher@gmail.com (C.I. Imakando).

https://doi.org/10.1016/j.agee.2023.108363

Received 17 November 2022; Received in revised form 28 December 2022; Accepted 16 January 2023 Available online 27 January 2023 0167-8809/© 2023 Elsevier B.V. All rights reserved. spend more time foraging in areas where they perceive the least fear from predation (Ylonen et al., 2002). Understanding how the foraging activity of rodents changes over the maize growing season could help to develop management strategies that incorporate the 'landscape of fear' (LOF) concept (Laundré et al., 2001) and thereby reduce rodent foraging in cropping areas. Furthermore, evidence-based information on where and when to conduct rodent management can help to prevent rodent outbreaks and is vital given that most farmers are resource poor and depend on agriculture for food and income (Swanepoel et al., 2017; Taylor et al., 2012).

To monitor and/or map the foraging activity of rodents across the maize growing season, one technique that can be exploited is the givingup-density (GUD) approach, which attempts to characterise the LOF for a species in a habitat. The LoF is "the spatially explicit distribution of perceived predation risk as seen by a prey population" (Bleicher, 2017; Gaynor et al., 2019). The perceived predation risk (cost of foraging) of a population can be measured by the GUD (Brown and Kotler, 2004). According to Johnson and Horn (2008), a forager abandons a patch quickly when the perceived risk of predation is high, leaving behind greater density of food compared to when the perceived risk of predation is low. In many crops, landscape features can affect both domestic and wild predators that prev on rodents (Pita et al., 2009; Fischer and Schröder, 2014; St. George and Johnson, 2021), as can the presence of farmers regularly tending their fields who may influence the spatial behaviour of rodents (Jones et al., 2017). Rodents use both direct (predator odours) and indirect (habitat type and weather conditions) cues to assess the risk of predation in a particular patch (Orrock et al., 2004).

GUDs have been successfully used to understand the foraging activity of rodents in rice fields (Jones et al., 2017), maize fields (Mohr et al., 2003), wheatfields (Ylonen et al., 2002) and in natural habitats (Wheeler and Hik, 2014; Yang et al., 2016; Loggins et al., 2019). Despite the application of GUD studies on rodents, few papers directly apply GUDs to assess rodent management strategies (Krijger et al., 2017). Currently, most rodent management strategies in maize cropping systems do not incorporate the rodent's landscape of fear which could increase their efficiency and reduce damage to maize crops. To address these shortcomings and highlight how GUD studies could refine management strategies beyond the usual measures of abundance or activity, the current study interprets the results with consideration of rodent pest management strategies by recommending areas (in and around maize fields) where farmers should focus pest control during different times of the maize cropping season, i.e., areas where rodents perceive the lowest levels of predation (Krijger et al., 2017). This is the first application of GUDs and LOF in Africa to understand how the foraging activity of rodent pest species in and around maize fields changes across a growing season.

We tested the hypothesis that the foraging activity of rodents in a maize cropping system is influenced by vegetation cover and food availability. We predicted that rodents will have: (i) lower foraging activity in the maize fields than along the border and in the adjacent natural habitat during the land preparation, planting, germination and post-harvest stages (when the maize fields have less vegetation cover and less food resources (grains)); (ii) equal foraging activity in maize fields and adjacent natural habitat from the weeding to maize tasselling stages when the vegetation cover in the maize fields increases; and (iii) the foraging activity will be higher in the maize fields than the adjacent habitat during the maturity, and harvest stages due to increased food resources (maize grains) and vegetation cover. This study will help to understand how different habitats may affect anti-predator and foraging activity and could guide rodent damage assessments (Jones et al., 2017) and guide future ecologically-based rodent management strategies (Krijger et al., 2017) in maize cropping systems.

2. Materials and methods

2.1. Experimental design

Four maize fields located adjacent to a natural habitat in Luto agricultural camp, Kitwe, Zambia (located between 12.94S,28.17E and 12.93S,28.20E) were selected for this study (Fig. 1). The maize fields ranged from 2 to 4 ha.

Prior to the main study, a pilot study was conducted to establish the best food (e.g., groundnuts, sunflower kernels, or pumpkin seeds), feeder (e.g., plastic, or wooden trays) and substrate type (sand or soil from the fields) for GUD estimates (Bedoya-Perez et al., 2013). The pilot study involved placing 20 seeds of a single type (groundnuts, sunflower kernels, or pumpkin seeds) in separate plastic or wooden trays (18 buried and 2 placed on the top) filled with either sand or soil from the fields. Trays contained small drainage holes in the bottom to allow rainwater to drain through. The trays were left in fields for three consecutive nights, counting the number of seeds remaining each morning, and resetting to 20 seeds each night. Pilot data indicated sunflower kernels and pumpkin seeds were more difficult to recover than groundnuts when counting the number of seeds remaining, especially if it had rained. Hence, we settled for groundnuts as the best food for the main study. Plastic trays were preferred over wooden trays because wooden trays were soaked by the rains making them difficult to carry around. Rodent activity was generally lower in trays filled with sand, possibly because the sand increased neophobic behaviour of rodents as sand is not commonly found in the area. Therefore, we used soils from the fields as the substrate type for the main study. Thus, the main study was developed using four plastic trays with 20 m spacing between the trays (Ylonen et al., 2002) which were placed along five transects. The first transect was laid along the border (field edge) of the maize field (transect #3), and two were at 20 m and 40 m either side of the border transect in maize fields (transects 1 and 2) and natural habitat (transects 4 and 5), respectively (Fig. 2).

Twenty groundnuts were placed in each tray, 18 buried in soil and 2 placed on the top (Jones et al., 2017). The foraging activity was monitored for three consecutive days (in the morning), with trays restocked with 20 groundnuts each day. The GUD was assessed by counting the number of seeds remaining in the tray (Brown, 1988). To confirm rodent activity at the feeding patches, a Bushnell Trophy Cam HD Essential Trail camera trap was placed in one patch per field at the beginning of the study (Fig. 3) (see Bedoya-Perez et al., 2013). The camera traps were set to record 24 h per day with a 30 s delay between detections (Williams et al., 2018; Rich et al., 2017). The following parameters were set on the camera traps; take three photos (8 M pixel) per trigger, sensor level at auto, NV shutter at medium and time stamp on. The camera traps were set 20 cm (Ramesh and Downs, 2015; Meek et al., 2012) above the ground on a wooden pole. The plastic tray was placed 1.5 m in front of the camera trap (Glen et al., 2016; Meek et al., 2012).

Foraging activity was assessed monthly (4-week intervals) during the maize growing season (October to July), while the populations of rodents in the maize fields were monitored monthly following the capturemark-recapture (CMR) procedure in fields 1-3 as part of another study which examined the population dynamics of small mammals in maize fields (Imakando, 2021). A 70 m \times 70 m permanent trapping grid was established in the centre of each maize field. Each trapping grid had seven trap lines, 10 m apart. Seven trapping stations, 10 m apart, were marked on each trap line. One Sherman live-trap, baited with a mixture of peanut butter and maize bran, was set in each trapping station. The traps were set in the evening and checked in the morning for three consecutive days in each grid. On the first capture, all animals were toe-clipped using sterile scissors. Rainfall data were accessed from the Copperbelt University meteorological station. This study was initially conducted during the 2018/2019 maize cropping season and repeated in the 2019/2020 maize cropping season.



Fig. 1. Map showing the location of the four maize fields in Luto agricultural camp, Kitwe, Copperbelt Province, Zambia.



Fig. 2. Layout of GUD trays in and around maize fields. The distance between the trays and transects was 20 m.

2.2. Statistical analysis

An independent samples t-test, with Levene's Test for equality of variance, was used to compare the GUD results from 2018/2019 and 2019/2020 cropping seasons. A two-way ANOVA using general linear mixed-effects model "package lme4" (Bates et al., 2019) was used to analyse the effect of distance (transect location) and crop stage on angular transformed GUDs (proportion of groundnuts remaining) (Laundré et al., 2001; Kasuya, 2004). Tukey's post hoc comparisons were conducted on distance (transects) and crop stage using the package 'multcomp' (Hothorn et al., 2019). All analyses were conducted in R version 3.6.1 (R Core Team, 2019). We used the minimum number of animals known to be alive (MNA) method to calculate the population of rodents during each stage.



Fig. 3. Camera trap image of *Mastomys natalensis* feeding from a tray used in assessing giving up density. Camera type (Bushnell Trophy Cam HD Essential Trail Camera) produces a 'black' infra-red flash that does not disturb mammal behaviour.

3. Results

3.1. Rodent species and population dynamics

From the CMR data, the most common rodent species in maize fields was *Mastomys natalensis* (87.05% of the total captures, Table 1).

The density of rodents was low during the planting period and increased as the vegetation increased in the maize fields. The highest rodent density in maize fields was observed during the harvest stage and just before the fields are cleared (May-June). Population density reduced during land preparation post-harvest, especially after clearing of the

Table 1

Species composition of small mammals (rodents and shrews) captured in three maize fields in Kitwe, Zambia, with species ordered by overall abundance. The numbers in brackets are percentage composition of each species.

| | Fields | | | |
|--------------------------|------------|------------|------------|-----------|
| Species | Luto 1 | Luto 2 | Luto 3 | Overall |
| Mastomys natalensis | 131 | 396 | 347 | 874 |
| | (72.78 %) | (91.45 %) | (88.75 %) | (87.05 %) |
| Mus minutoides | 23 (12.78 | 15 (3.46 | 18 (4.60 | 56 (5.58 |
| | %) | %) | %) | %) |
| Crocidura hirta | 14 (7.78 | 6 (1.39 %) | 11 (2.05 | 28 (2.79 |
| | %) | | %) | %) |
| Steatomys pratensis | 2 (1.11 %) | 3 (0.69 %) | 11 (2.81 | 16 (1.59 |
| | | | %) | %) |
| Saccostomus campestris | 5 (2.78 %) | 7 (1.62 %) | 1 (0.26 %) | 13 (1.29 |
| | | | | %) |
| Gerbilliscus leucogaster | 3 (1.67 %) | 6 (1.67 %) | 1 (0.26 %) | 10 (1.00 |
| | | | | %) |
| Elephantulus | 0 (0 %) | 0 (0 %) | 2 (0.51 %) | 2 (0.2 %) |
| brachyrhynchus | | | | |
| Rattus rattus | 1 (0.56 %) | 0 (0 %) | 1 (0.26 %) | 2 (0.2 %) |
| Lemniscomys rosalia | 0 (0 %) | 0 (0 %) | 1 (0.26 %) | 1 (0.1 %) |
| Acomys spinosissimus | 0 (0 %) | 0 (0 %) | 1 (0.26 %) | 1 (0.1 %) |
| Arvicanthis niloticus | 1 (0.56 %) | 0 (0 %) | 0 (0 %) | 1 (0.1 %) |
| Total | 180 (100 | 433 (100 | 391 (100 | 1004 (100 |
| | %) | %) | %) | %) |
| Species richness | 8 | 6 | 10 | 11 |
| Shannon-weaver | 0.97 | 0.42 | 0.56 | 0.56 |
| Diversity index | | | | |

fields (Fig. 4).

3.2. Changes in the foraging activity of rodent pest species in and around maize fields

Motion sensitive camera traps confirmed that *M. natalensis* was the rodent species that frequently visited the GUDs. An independent samples t-test was conducted to examine GUD differences between the 2018/2019 and 2019/2022 cropping seasons. Levene's Test for equality of variances showed no violations, p = 0.289. In general, the GUD results from the 2018/2019 season and 2019/2020 season were not significant different (t(88) = 1.201, p = 0.233), so the data were combined during the analyses. Due to differences in the planting and harvest times between farmers, the results from December and January were combined as "germination stage" while the results from June and July were combined as "post-harvest" during the analyses, but these were separated when constructing a heat map on spatial use. In general, the foraging activity of rodents was highest (i.e., lowest GUD) during the

germination stage (mean GUD = 50.5) while lowest during the land preparation and post-harvest stages (mean GUD = 58.0 at both crop stages). From the two-way ANOVA, overall, there was a significant effect of distance for crop stage ($F_{7,40} = 11.228, p < 0.001$) and the interaction between the effects of distance and crop stage on the GUD ($F_{28,40} = 4.723, p < 0.001$), but no effect of distance on the GUD ($F_{4,40} = 1.631, p = 0.185$).

For distance from the edge of crops and natural habitat (transect), Tukey's post-hoc tests revealed that rodent activity was affected during maize germination and harvest stages, whereas rodent activity was similar on all transects during the other stages (see Supplementary Table S2). During the maize germination stage, rodent activity was higher on transect 5 (40 m into the natural habitat; mean GUD = 37) than on transects 1 (mean GUD = 59; t = -6.551, p < 0.001), 2 (mean GUD = 58; t = -6.039, p < 0.001) and 3 (mean GUD = 53; t = -3.941, p = 0.001). Additionally, rodent activity was higher on transect 4 (20 m into the natural forest; mean GUD = 43) than on transects 1 (t = -4.528, p < 0.001), and 2 (t = -4.016, p < 0.001) during the germination stage. However, rodent activity during the harvest stage was higher on transect 1 (40 m into the maize field; mean GUD = 50) than on transects 3 (mean GUD = 59; t = 2.935, p = 0.034), 4 (mean GUD = 60; t = 3.277, p = 0.013) and 5 (mean GUD = 59; t = 2.817, p = 0.047).

Rodent activity was higher during the germination stage than during land preparation, planting, weeding, maturation, harvest and post-harvest crop stages [(LP vs. G; z = -5.690, p < 0.001); (P vs. G; z = -4.757, p < 0.001); (G vs. W; z = 3.401, p = 0.015); (G vs. M; z = 3.409, p = 0.015); (G vs. H; z = 5.418, p < 0.001); and (G vs. PH; z = 6.757, p < 0.001)]. Rodent activity also was higher during the maize tasselling stage than land preparation (z = -3.154, p = 0.034); and higher during the weeding stage than harvest stage (z = 3.262, p = 0.024) (see Supplementary Table S3). The changes in the foraging activity of rodents in and around maize fields can be summarised using a heat map (Fig. 5).

4. Discussion

This is the first study to document how the foraging activity of rodents in and around maize fields changes across the growing season. As predicted, the foraging activity of rodents over the maize growing season was influenced by vegetation cover and food availability. Higher rodent activity occurred in the adjacent natural habitat than along and inside the maize field during the germination period. Uniform/equal rodent activity in the adjacent natural habitat, along the border and inside maize fields occurred during the land preparation, planting, weeding, maize tasselling, maturity, and post-harvest stages. During the harvesting period rodent activity was significantly higher inside the



Fig. 4. Mean monthly rodent abundance (minimum number known to be alive) in maize fields (n = 3) in Kitwe, Zambia. The letters below the months represents seasons; WW, warm-wet season (November to April); CD, cold-dry season (April to August); HD, hot-dry season (September to October).



Fig. 5. Heat map showing rodent foraging activity across the maize growing season. The lower the mean GUD, the higher the rodent foraging activity and vice versa. The letters on the x-axis represent crop stage; LP = land preparation (October); P = planting stage (November); G = germination stages (December and January); W = weeding stage (February); MT = maize tasselling stage (March); M = maturity stage (April); H = harvesting stage (May); and PH = post-harvest stage (June and July).

maize fields than along the border and adjacent natural habitat. These results provide evidence-based information on how rodent foraging activity changes across the maize growing season. Krijger et al. (2017) suggested that focusing rodent pest management in those areas where rodents perceived the lowest predation risk could be more effective and efficient. Therefore, in management terms, our findings suggest that farmers are likely to have greater success managing rodent pests during the germination of the maize crop by focusing rodent control measures in the natural habitats adjacent to the maize fields rather than inside or along the edge of maize fields. From the weeding to maturation stages, equal success would be achieved if rodent control measures are concentrated inside, along the edge of maize fields or in the natural habitat adjacent to the maize fields. During the harvest stage, greater success in managing rodent pests could be achieved if rodent control measures are concentrated inside the maize fields than if rodent control measures are concentrated along the field border or in the natural habitat adjacent to the maize fields. However, it is necessary to survey the species composition in all the habitats before application of control measures to minimise non-target implications of rodent control in natural habitats. Therefore, our findings can be used by small holder farmers to focus rodent management strategies efficiently and effectively at different stages in the maize cropping season, which in turn will reduce the cost for controlling rodent pest species and losses of the crops.

Earlier work in maize fields in Tanzania have shown that vegetation cover plays an important role in the foraging, habitat preference and population dynamics of rodents, particularly M. natalensis (Leirs et al., 1996; Mohr et al., 2003). Leirs et al. (1996) reported that M. natalensis preferred (i.e., was more active in) areas with vegetation cover while tending to avoid open spaces, especially during periods of low density. Corroborating this finding, Mohr et al. (2003) used GUD and video evidence to show that M. natalensis perceived lower predation risk in feeding patches with cover than in open patches. Elsewhere, research on the foraging activity of other rodents using GUDs suggests that rodent foraging activity is shaped by the perceived predation risk (Ylonen et al., 2002; Orrock et al., 2004; Wheeler and Hik, 2014; Jones et al., 2017). Jones et al. (2017) found that the foraging activity of Rattus tanezumi in rice fields in the Philippines was shaped by the perceived predation risk whereby more damage was observed in the middle of the rice fields (with more vegetation cover) than on the borders and rice bund, with no vegetation. Ylonen et al. (2002) reported that prior to harvest of wheat in southern Australia, house mice, Mus domesticus, were mainly in the crop. Similarly, Oldfield mice, Peromyscus polionotus, in South Carolina, USA, were found to remove more seeds in areas with vegetation cover than outside of cover (Orrock et al., 2004). In Canada, the arctic ground

squirrel, *Urocitellus parryii*, exhibited habitat specific strategies to minimise predation risk by foraging more in tundra and shrub-tundra habitats while avoiding the shrub-dominated habitat, which reduced their visibility and increased predation risk (Wheeler and Hik, 2014). All these studies highlight the importance of vegetation cover as a feature of small mammal behaviour to avoid predation, which aligns with the conclusions from our study.

Increased rodent activity around maize fields during the germination stage was reported in other studies (Stenseth et al., 2003; Mulungu et al., 2007). However, the findings in our study indicate that rodent activity was only high in the adjacent natural habitat and along the border during germination of the maize crop. This indicates that, during the germination stage, the perceived predation risk was higher inside the maize fields than along the border and adjacent natural habitat, corroborating the studies by Johnson and Horn (2008) and Jones et al. (2017), who reported that rodents perceived open areas to be riskier than areas with cover. Similarly, Key (1990) found that pre-harvest maize damage from the African ground squirrel occurred at the edges of the fields than in the middle and that they used the edges of the field for refuge when disturbed while feeding in the fields. However, when farming methods that lead to less disturbance to rodent burrows and increased cover and food supply, such as conservation agriculture, mice become resident in fields all the time, rather than retreating to field edges (Ruscoe et al., 2022). This indicates that foraging activity of rodents in maize fields is shaped by their perceived predation risk. Therefore, high GUDs during the land preparation stage on all transects may be because at this stage the maize fields were cleared and, even in the natural habitat, the vegetation cover is dry and minimal.

As the height of maize and vegetation increased inside the maize fields (from the weeding to maturation stages), there was no difference in the mean GUD between the natural habitat and maize field transects indicating that the perceived predation was equal in the forest, along the margin and inside the maize field. This further supports the contention that rodent foraging activity is shaped by vegetation cover (Brown, 1988; Mohr et al., 2003). Vegetative cover provides shelter for rodents, leading to reduced detection probability and capture by predators (Banasiak and Shrader, 2016) and thus reduces the perceived predation risk and increases the foraging activity of rodent species (Loggins et al., 2019). A limitation of our study is that we are unable to comment on whether the different rodent species found in the study area respond to vegetation cover in the same way. Further studies, for example using camera traps, are recommended to understand whether there are detectible differences in the way different small mammal species within the same habitat respond to GUDs, vegetation cover and predation risk.

As our study area is dominated by *M. natalensis*, we can expect our results closely align with the behaviour of this species.

Increased rodent activity inside the maize fields compared to along the border and adjacent habitat during the harvest stage suggests that, at this stage, the foraging activity was shaped by both vegetation and increased availability of food resources (Sluydts et al., 2007). In addition, the harvest stage (May) coincided with the peak rodent population (see Fig. 4) in maize fields. When presented with patches of equal vegetation cover, food availability becomes important in explaining the foraging activity of rodents. This finding supports the hypothesis that rodents select to forage in habitats and microhabitats where the perceived risk of predation is low (Brown, 1988; Jacob and Brown, 2000; Ylonen et al., 2002). These findings support the suggestion that rodent management during the harvest stage would be more successful by placing baits inside the maize field than along the border or adjacent natural habitat.

In conclusion, GUD was successfully used to monitor rodent foraging activity in and around maize fields over the maize cropping season. Rodent activity was driven by vegetation cover and food availability. Based on our findings, during the germination period, rodent control measures should be concentrated along the maize fields edges and in the natural habitat adjacent to maize fields while during the harvest period rodent control measures should be concentrated inside the maize fields. This information will help smallholder farmers to be more efficient and effective in rodent control by focusing their management strategies in areas of perceived reduced predation risk (Krijger et al., 2017). We recommend further research, such as using rodenticide baits or methods of trapping (e.g., linear trap barrier) at different times of the growing season and at different distances from the maize field, to assess the effect on rodent population dynamics and associated crop losses. Follow up studies should also collect and include data on plant biomass on field edges and within fields to track the changes over time and then assess how these changes affect rodent foraging activity. Additionally, farming practices such as tractor ploughing and management of the vegetation around fields margins can be used to increase predation risk (Brown et al., 2004: Massawe et al., 2006). Target rodent pest species and non-target impacts should be surveyed in adjacent natural habitats, particularly as these habitats provide a range of ecosystem services beneficial to agricultural production (Hatt et al., 2017; Lindell et al., 2018; Mkenda et al., 2019) where trade-offs between the management of rodents, insects, and weeds as well as crop pollination services need careful cost-benefit assessments (Wegner and Pascual, 2011; Wratten et al., 2012; Williams et al., 2018).

Funding

This research was supported by the Commonwealth Scholarship Commission, UK with partial funding from the African Union (grant # AURG II-1-006–2016 EcoRodMan).

CRediT authorship contribution statement

ICI and SRB conceived the topic. ICI, SRB, MFG and GRS conceived the research. ICI conducted the field work and analyzed the data. All the authors contributed to the writing of the manuscript.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

Acknowledgements

We thank Remmy Kopeka, Oliver Susiku, Mabvuto Phiri and Fabian Chulu for their valuable assistance during the field work. We thank Darius Phiri for the help in designing the study Map. We thank the farmers of Luto agricultural camp for hosting the project. We also thank DNPW for the research permit. We thank the Ministry of Agriculture (Kitwe Office) for facilitating collaboration with the farmers. Ethical clearance to carry out the work was granted by the University Research Ethics Committees of the University of Greenwich (UREC/17.2.5.13) and the Copperbelt University.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.agee.2023.108363.

References

- Arnold, S.E.J., Elisante, F., Mkenda, P.A., Tembo, Y.L.B., Ndakidemi, P.A., Gurr, G.M., Darbyshire, I.A., Belmain, S.R., Stevenson, P.C., 2021. Beneficial insects are associated with botanically rich margins with trees on small farms. Sci. Rep. 11, 1–11.
- Banasiak, N., Shrader, A.M., 2016. Similarities in perceived predation risk prevent temporal partitioning of food by rodents in an African grassland. J. Mammal. 97, 483–489.
- Bates, D., Maechler, M., Bolker, B., Walker, S., Christensen, R.H.B., Singmann, H., Dai, B., Scheipl, F., Grothendieck, G., Green, P., John, F. (2019) Package 'lme4': linear mixed-effects models using 'Eigen' and S4. Version 1.1-21. Version 1., 1–123.
- Bedoya-Perez, M.A., Carthey, A.J.R., Mella, V.S.A., McArthur, C., Banks, P.B., 2013. A practical guide to avoid giving up on giving-up densities. Behav. Ecol. Sociobiol. 67, 1541–1553.
- Belmain, S.R., 2010. Developing pesticide-free rodent control for southern Africa. Pestic. N. 87, 9–11.
- Bleicher, S.S., 2017. The landscape of fear conceptual framework: definition and review of current applications and misuses. PeerJ 5, e3772.
- Brown, J.S., 1988. Patch use as an indicator of habitat preference, predation risk, and competition. Behav. Ecol. Sociobiol. 22, 37–47.
- Brown, J.S., Kotler, B.P., 2004. Hazardous duty pay and the foraging cost of predation. Ecol. Lett. 7, 999–1014.
- Brown, P.R., Davies, M.J., Croft, J.D., Singleton, G.R., 2004. Can farm management practices reduce the impact of house mouse populations on crops in an irrigated farming system? Wildl. Res. 31, 597–604.
- Fischer, C., Schröder, B., 2014. Predicting spatial and temporal habitat use of rodents in a highly intensive agricultural area. Agric. Ecosyst. Environ. 189, 145–153.
- Gaynor, K.M., Brown, J.S., Middleton, A.D., Power, M.E., Brashares, J.S., 2019. Landscapes of fear: spatial patterns of risk perception and response. Trends Ecol. Evol. 34, 355–368.
- Glen, A., Anderson, D., Veltman, C., Garvey, P., Nichols, M., 2016. Wildlife detector dogs and camera traps: a comparison of techniques for detecting feral cats. N. Z. J. Zool. 43, 127–137.
- Hatt, S., Lopes, T., Boeraeve, F., Chen, J., Francis, F., 2017. Pest regulation and support of natural enemies in agriculture: experimental evidence of within field wildflower strips. Ecol. Eng. 98, 240–245.
- Hothorn, Bretz, F., Westfall, P., Heiberger, R.M., Schuetzenmeister, A., Scheibe, S. (2019) Multcomp package (Simultaneous Inference in General Parametric Models). https:// cran.r-project.org/web/packages/multcomp/multcomp.pdf.
- Imakando, C.I., 2021. Implications of Habitat Fragmentation on Small Diversity, Rodent Pest Regulation and Ecologically Based Management Strategies in Kitwe, Zambia. University of Greenwich.
- Jacob, J., 2008. Response of small rodents to manipulations of vegetation height in agroecosystems. Integr. Zool. 3, 3–10.
- Jacob, J., Brown, J.S., 2000. Microhabitat use, giving-up densities and temporal activity as short-and long-term anti-predator behaviors in common voles. Oikos 91, 131–138.
- Johnson, M.D., Horn, C.M., 2008. Effects of rotational grazing on rodents and raptors in a coastal grassland. West. North Am. Nat. 68, 444–452.
- Jones, C.R., Lorica, M.R.P., Villegas, J.M., Ramal, A.F., Horgan, F.G., Singleton, G.R., Stuart, A.M., 2017. The stadium effect: rodent damage patterns in rice fields explored using giving-up densities. Integr. Zool. 12, 438–445.
- Kasuya, E., 2004. Angular transformation another effect of different sample sizes. Ecol. Res. 19, 165–167.
- Key, G., 1990. Pre-harvest crop losses to the African striped ground squirrel, Xerus erythropus in Kenya. Trop. Pest Manag. 36, 223–229.
- Krijger, I.M., Belmain, S.R., Singleton, G.R., Groot Koerkamp, P.W., Meerburg, B.G., 2017. The need to implement the landscape of fear within rodent pest management strategies. Pest Manag. Sci. 73, 2397–2402.
- Laundré, J.W., Hernández, L., Altendorf, K.B., 2001. Wolves, elk, and bison: reestablishing the 'landscape of fear' in Yellowstone National Park, U.S.A. Can. J. Zool. 79, 1401–1409.

C.I. Imakando et al.

- Leirs, H., Verheyen, W., Verhagen, R., 1996. Spatial patterns in Mastomys natalensis in Tanzania (Rodentia, Muridae). Mammalia 60, 545–555.
- Lindell, C., Eaton, R.A., Howard, P.H., Roels, S.M., Shave, M.E., 2018. Enhancing agricultural landscapes to increase crop pest reduction by vertebrates. Agric., Ecosyst. Environ. 257, 1–11.
- Loggins, A.A., Shrader, A.M., Monadjem, A., McCleery, R.A., 2019. Shrub cover homogenizes small mammals' activity and perceived predation risk. Sci. Rep. 9, 1–11.
- Massawe, A.W., Rwamugira, W., Leirs, H., Makundi, R.H., Mulungu, L.S., 2006. Do farming practices influence population dynamics of rodents? A case study of the multimammate field rats, Mastomys natalensis, in Tanzania. Afr. J. Ecol. 45, 293–301.
- Meek, P.D., Ballard, G., Fleming, P., 2012. An introduction to camera trapping for wildlife surveys in Australia. Canberra, Austalia: PestSmart Toolkit publication, Invasive Animals Cooperative Research Centre.
- Mkenda, P.A., Ndakidemi, P.A., Mbega, E., Stevenson, P.C., Arnold, S.E.J., Gurr, G.M., Belmain, S.R., 2019. Multiple ecosystem services from field margin vegetation for ecological sustainability in agriculture: Scientific evidence and knowledge gaps. PeerJ 7 (e8091), 1–33.
- Mohr, K., Vibe-Petersen, S., Jeppesen, L.L., Bildsøe, M., Leirs, H., 2003. Foraging of multimammate mice, *Mastomys natalensis*, under different predation pressure: cover, patch-dependent decisions and density-dependent GUDs. Oikos 100, 459–468.
- Mulungu, L.S., 2017. Control of rodent pests in maize cultivation: the case of Africa. In: Watson, D. (Ed.), Achieving Sustainable Maize Cultivation. Burleigh Dodds Science Publishing Limited, pp. 317–338.
- Mulungu, L.S., Makundi, R.H., Massawe, A.W., Machang'u, R.S., Ngowo, V., Leirs, H., 2005. Spatial patterns and distribution of damage in maize fields due to *Mastomys natalensis* in Tanzania. Belg. J. Zool. 135 (supplement), 183–185.
- Mulungu, L.S., Makundi, R.H., Massawe, A.W., Leirs, H., 2007. Relationship between sampling intensity and precision for estimating damage to maize caused by rodents. Integr. Zool. 2, 131–135.
- Ochieng, L.O., Ogendo, J.O., Bett, P.K., Nyaanga, J.G., Cheruiyot, E.K., Mulwa, R.M.S., Arnold, S.E.J., Belmain, S.R., Stevenson, P.C., 2022. Field margins and botanical insecticides enhance *Lablab purpureus* yield by reducing aphid pests and supporting natural enemies. J. Appl. Entomol. 146, 838–849.
- Orrock, J.L., Danielson, B.J., Brinkerhoff, R.J., 2004. Rodent foraging is affected by indirect, but not by direct, cues of predation risk. Behav. Ecol. 15, 433–437.
- Pita, R., Mira, A., Moreira, F., Morgado, R., Beja, P., 2009. Influence of landscape characteristics on carnivore diversity and abundance in Mediterranean farmland. Agric., Ecosyst. Environ. 132, 57–65.
- R Core Team, 2019. R: A language and environment for statistical computing. R foundation for statistical computing,, Vienna, Austria (URL). (https://www.R-proje ct.org/).
- Ramesh, T., Downs, C.T., 2015. Impact of land use on occupancy and abundance of terrestrial mammals in the Drakensberg Midlands, South Africa. J. Nat. Conserv. 23, 9–18.
- Rich, L.N., Miller, D.A.W., Robinson, H.S., McNutt, J.W., Kelly, M.J., 2017. Carnivore distributions in Botswana are shaped by resource availability and intraguild species. J. Zool. 303, 90–98.

- Rodríguez-Pastor, R., Luque-Larena, J.J., Lambin, X., Mougeot, F., 2016. "Living on the edge": The role of field margins for common vole (*Microtus arvalis*) populations in recently colonised Mediterranean farmland. Agric., Ecosyst. Environ. 231, 206–217.
- Ruscoe, W.A., Brown, P.R., Henry, S., van de Weyer, N., Robinson, F., Hinds, L.A., Singleton, G.R., 2022. Conservation agriculture practices have changed habitat use by rodent pests: implications for management of feral house mice. J. Pest Sci. 95, 493–503.
- Skonhoft, A., Leirs, H., Andreassen, H.P., Mulungu, L.S.A., Nils, C.S., 2006. The bioeconomics of controlling an African rodent pest species. Environ. Dev. Econ. 11, 453–475.
- Sluydts, V., Crespin, L., Davis, S., Lima, M., Leirs, H., 2007. Survival and maturation rates of the African rodent, *Mastomys natalensis*: density-dependence and rainfall. Integr. Zool. 2, 220–232.
- St. George, D.A., Johnson, M.D., 2021. Effects of habitat on prey delivery rate and prey species composition of breeding barn owls in winegrape vineyards. Agric. Ecosyst. Environ. 312, 107322.
- Stenseth, N.C., Leirs, H., Skonhoft, A., Davis, S.A., Pech, R.P., Andreassen, H.P., Singleton, G.R., Lima, M., Machang'u, R.S., Makundi, R.H., Zhang, Z., Brown, P.R., Shi, D., Wan, X., 2003. Mice, rats, and people: the bio-economics of agricultural rodent pests. Front. Ecol. Environ. 1, 367–375.
- Swanepoel, L.H., Swanepoel, C.M., Brown, P.R., Eiseb, S.J., Goodman, S.M., Keith, M., Kirsten, F., Leirs, H., Mahlaba, T.A.M., Makundi, R.H., Malebane, P., Von Maltitz, E. F., Massawe, A.W., Monadjem, A., Mulungu, L.S., Singleton, G.R., Taylor, P.J., Soarimalala, V., Belmain, S.R., 2017. A systematic review of rodent pest research in Afro-Malagasy small-holder farming systems: are we asking the right questions? PLOS One 12, 1–20.
- Taylor, P.J., Downs, S., Monadjem, A., Eiseb, S.J., Mulungu, L.S., Massawe, A.W., Mahlaba, T.A., Kirsten, F., Von Maltitz, E., Malebane, P., Makundi, R.H., Lamb, J., Belmain, S.R., 2012. Experimental treatment-control studies of ecologically based rodent management in Africa: balancing conservation and pest management. Wildl. Res. 39, 51–61.
- Wegner, G., Pascual, U., 2011. Cost-benefit analysis in the context of ecosystem services for human well-being: a multidisciplinary critique. Glob. Environ. Change 21, 492–504.
- Wheeler, H.C., Hik, D.S., 2014. Giving-up densities and foraging behaviour indicate possible effects of shrub encroachment on arctic ground squirrels. Anim. Behav. 95, 1–8.
- Williams, S.T., Maree, N., Taylor, P., Belmain, S.R., Keith, M., Swanepoel, L.H., 2018. Predation by small mammalian carnivores in rural agro-ecosystems: an undervalued ecosystem service? Ecosyst. Serv. 30, 362–371.
- Wratten, S.D., Gillespie, M., Decourtye, A., Mader, E., Desneux, N., 2012. Pollinator habitat enhancement: Benefits to other ecosystem services. Agric. Ecosyst. Environ. 159, 112–122.
- Yang, Y., Zhang, M., Yi, X., 2016. Small rodents trading off forest gaps for scatterhoarding differs between seed species. For. Ecol. Manag. 379, 226–231.
- Ylonen, H., Jacob, J., Davies, M.J., Singleton, G.R., 2002. Predation risk and habitat selection of Australian house mice, *Mus domesticus*, during an incipient plague: desperate behaviour due to food depletion. Oikos 99, 284–289.